RESEARCH ARTICLE



# Exercise training in ad libitum and food-restricted old rats: effects on metabolic and physiological parameters

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Received: 2 August 2019 / Accepted: 14 October 2019 © Springer Nature B.V. 2019

Abstract Aging is accompanied by a decline in the healthy function of multiple organs, leading to increased incidence and mortality from diseases such as cancer and inflammatory, cardiovascular and neurodegenerative diseases. Dietary restriction is the most effective experimental intervention known to consistently slow the aging process and with positive effects on health span in different organisms, from invertebrates to mammals. Age is also associated with progressive decline in physical activity levels in a wide range of animal species: therefore, regular physical exercise could represent a safe intervention to antagonize aging. In this research we explore the effects of exercise training initiated in late middle aged rats fed with different lifelong dietary regimens: one group was fed ad libitum and the second group was subjected to every-other-day fasting. These two groups might represent examples of ''normal'' aging

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and ''successful'' aging. The study shows the effects of exercise and food restriction and their interaction on plasma levels of total antioxidant capacity, lactate, amino acids, and on products of protein oxidation in soleus and tibialis anterior muscles. In addition, we evaluated body composition measurement by bioelectrical impedance analysis and muscle strength by grasping test. Results show that late-onset exercise training has the potential to improve some metabolic and physiological parameters in rats with the same ''chronological age'' but different ''biological age'', without negative effects, and highlight the relevance of a personalised and selected exercise protocol, since the responsiveness to exercise may depend on the individual's ''biological age''.

Keywords Aging · Dietary restriction · Exercise · Rat

### Introduction

Aging is characterized by a complex interaction of stochastic, environmental, genetic and epigenetic variables. This interaction generates the loss of molecular accuracy and therefore a random accumulation of damage in the organism's cells, tissues, or whole organism during life increases: the probability of disease and death also augments in proportion

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(Rattan [2015](#page-12-0)). Indeed, aging is accompanied by a decline in the healthy function of multiple organs, leading to increased incidence and mortality from diseases such as cancer and inflammatory, cardiovascular and neurodegenerative diseases (Kennedy et al. [2014\)](#page-12-0). In recent years, there has been increasing interest in interventions to develop realistic and practical methods for maintaining health throughout the life span. DR is the most effective experimental intervention known to consistently slow the aging process and extend median and maximum life span with positive effects on health span in different organisms, from invertebrates to mammals (Kennedy et al. [2007](#page-12-0)). Observational studies suggest that DR may have beneficial effects also on human longevity (Heilbronn et al. [2006;](#page-12-0) Lefevre et al. [2009\)](#page-12-0). Recent studies indicate that the benefits of DR on aging are conserved in non-human primates (Mattison et al. [2017\)](#page-12-0). Age is also associated with progressive decline in physical activity levels in a wide range of animal species, ranging from the Caenorhabditis elegans worm (Herndon et al. [2002\)](#page-12-0) to humans (Westerterp [2015\)](#page-13-0), with major metabolic consequences (Chow et al. [2007;](#page-11-0) Westerterp [2013](#page-13-0)): therefore, regular physical exercise could represent a safe intervention to delay aging (Cobley et al. [2015](#page-11-0)). The available data strongly indicate that regular exercise plays a preventive role against lifestyle-dependent diseases (Radak et al. [2004;](#page-12-0) Goto and Radák [2009](#page-12-0)) and increase mean life span in rodents (Holloszy et al. [1985\)](#page-12-0). On the other hand, the well-documented beneficial effects of exercise occur in a paradoxical background of biochemical framework: it is well known that exercise increases the production of potentially harmful substances such as reactive oxygen and nitrogen species, other free radicals, acids and aldehydes (Alessio et al. [2000](#page-11-0); Sahlin et al. [2010](#page-12-0); Powers et al. [2016\)](#page-12-0). The paradox arises as to whether exercise would be recommended to aged population since senescent organisms may be more susceptible to increase of potentially harmful substances during exercise.

In order to obtain further evidence that can gain knowledge about the relationship between exercise and aging, in this research we explore the effects of exercise training initiated in late middle aged rats fed with different lifelong dietary regimens: one group was fed ad libitum (AL) and the second group was subjected to every-other-day fasting (EOD). These two groups might represent examples of ''normal''

aging and ''successful'' aging: AL rats are well fed laboratory animals as humans living in affluent western societies, and therefore they may be a model of ''normal'' aging; EOD rats are laboratory animals submitted to DR, the most effective experimental intervention known to consistently slow aging, and for this reason they may represent subjects who have taken measures to achieve a healthy and ''successful'' aging. The study shows the effects of exercise and food restriction and their interaction on plasma levels of total antioxidant capacity (TAC), lactate, AAs, and on products of protein oxidation in soleus and tibialis anterior muscles. In addition, we evaluated the effects of treatments on physiological parameters: body composition measurement by BIA and muscle strength by grasping test. The metabolic and physiological dataset obtained from rats with the same ''chronological age'' but different ''biological age'' might be useful to understand whether exercise training, initiated in late middle age, may improve physical functions, and consequently quality of life of the elderly population.

### Materials and methods

### Materials

All reagents were of analytical and HPLC grade. Solvents were purchased from Panreac Química S.L.U. (Barcelona, Spain). Standard molecules and chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA). Milli-Q (Millipore-Lab, Bedford, MA, USA) purified water was used for all analyses.

#### Animals

Male Sprague–Dawley rats, raised in the Pisa University Interdepartmental Research Centre on Biology and Pathology of Aging Vivarium, were used. All procedures and animal treatment followed the European Community Directive 2010/63/UE and Italian animal welfare laws, guidelines, and policies. All handing and management procedures were approved by the Independent Ethics Committee of the University of Pisa (Approval number: No. 2A/4155).

Animals were kept in a controlled environment (22 °C, 12/12 h light/dark cycle), had free access to water, and fed AL with standard rodent diet (Teklad, Harlan, Italy) until 2 months of age. At that time, rats were randomly assigned to dietary treatments: one group was fed AL, the second group was subjected to EOD. At 19 months of age, AL and EOD rats were further divided in two sub-groups as: AL sedentary (ALs), AL exercised (ALe), EOD sedentary (EODs), and EOD exercised (EODe).

At 24 months of age, rats were sacrificed under pentobarbital anesthesia (50 mg/kg body weight, i.p.). Food was withdrawn 16 h before experimentation. The age of 24 months for sacrifice was chosen, because it represents approximatively the mean life span for Sprague–Dawley rats fed ad libitum (Masoro [1980\)](#page-12-0).

Blood was collected from the posterior vena cava into test tube containing 0.25 M EDTA, centrifuged, and plasma samples were stored at  $-80$  °C until analysis. Soleus (slow-twitch muscle, composed predominantly of red fibers) and tibialis anterior (fasttwitch muscle, composed predominantly of white fibers) muscles were dissected out, snap frozen in liquid nitrogen, and stored at  $-80$  °C until analysis.

### Exercise training programme

The exercise protocol was designed in accordance with the basic principles of training in humans: specificity, progressive overload, and variable intensity (Spiering et al. [2008](#page-13-0); Goutianos et al. [2015\)](#page-12-0). A strict control of health and animal welfare before, during, and after each training session was performed. Rats were not daily trained, but only three times a week to minimize the potentially evoked stress effects of exercise and to allow the recovery of liver and muscle glycogen.

Exercise was performed using a converted human treadmill with 5 separate running lanes with an incline of  $0^\circ$ ; an acrylic block was placed on the ledge at the end of the belt so as to make it difficult for rats to remain there. All rats assigned to trained groups were adapted to walking on a treadmill (2 weeks: three times a week for 20 min at 4 m/min), before the beginning of the exercise protocol. Exercise training programme was composed of two phases with sessions of 30 min three times a week. Training phase 1 (8 weeks) was set with a gradually increasing running speed and time, as follows: weeks 1–2 (4 min at 7 m/ min, 6 min at 9 m/min, 10 min at 12 m/min, 6 min at 9 m/min, 4 min at 7 m/min); weeks 3–4 (5 min at

7 m/min, 20 min at 12 m/min, 5 min at 7 m/min); weeks 5–6 (5 min at 9 m/min, 20 min at 15 m/min, 5 min at 9 m/min); weeks 7–8 (5 min at 12 m/min, 20 min at 15 m/min, 5 min at 12 m/min) (Ben et al. [2009,](#page-11-0) [2010\)](#page-11-0). Training phase 2 (2 weeks) was set with the running speed returned to the level at the beginning of phase 1 to reach adaptation at a stable training load. Training phases 1 and 2 were repeated to complete 5 months of exercise. When necessary for rats to run, their tails were stimulated using a soft bristle brush. To control for the stress of handling and exposure to the treadmill, sedentary animals (ALs and EODs groups) were placed on the stationary treadmill three times a week, 5 min per session, during the length of the study.

# Body composition measurement by bioelectrical impedance analysis

The effects of treatments on body composition in fat free mass and fat were measured by BIA. BIA procedure was conducted as described by Skalicky et al. [\(2001](#page-13-0)) at the beginning and at the end of exercise training programme. A tetrapolar impedance (model 101 RJL, Clinton T., MI) was used. Three consecutive measurements were performed. The equation validated against chemical carcass analysis by Skalicky et al. [\(2001](#page-13-0)) was used for the calculation of fat free mass and fat (in grams).

#### Grasping test

The grasping test is a simple non-invasive method designed to evaluate rodent forelimb muscle strength in vivo by taking advantage of the animal's tendency to grasp a horizontal metal bar (Smith et al. [1995\)](#page-13-0). The rat was placed over a base plate in front of a grasping bar, and to perform the evaluation the animal was pulled by the tail with increasing force. The rat could seize a grid attached to a force transducer until the animal lost its grip. The bar was attached to a force transducer (Ugo Basile Grip-Strength Meter), and the force produced during the pull on the bar was measured three times during each test. The grasping test was performed at the beginning and at the end of exercise training programme by the same investigator. Results are given as quotient between grip strength and animal body weight.

### Plasma analysis

#### Measurement of total antioxidant capacity

TAC assay considers the cumulative action of all the antioxidants present in plasma and may help in the measurement of physiological, environmental, and nutritional factors of the redox status. TAC was measured by the method based on the absorbance of the stable, colored 2,2'-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid radical cation)  $(ABTS^+)$ , as described by Erel [\(2004](#page-11-0)). The reduced ABTS molecule is oxidized to  $ABTS^+$  using hydrogen peroxide alone in acetate buffer (30 mmol/L, pH 3.6) and in this solution, the concentrate (deep green)  $ABTS^+$  molecules stay stable for a long time. When it is diluted with a more concentrated acetate buffer at high pH values (400 mmol/L, pH 5.8), the color is spontaneously and slowly bleached. Antioxidants present in the sample accelerate the bleaching rate to a degree proportional to their concentrations. The bleaching rate is inversely related with the TAC of the sample and this reaction can be monitored spectrophotometrically. The reaction rate was calibrated with Trolox, which is widely used as a traditional standard for TAC measurements assays. Results are given as mmol Trolox equivalent/L.

### Determination of lactate threshold

Before sacrifice, all rats performed an incremental test (IT) for evaluation of blood lactate threshold (LT) to verify the effect of treatments on physical capacity as described by Pilis et al. [\(1993](#page-12-0)) and Carvalho et al. [\(2005](#page-11-0)) with modifications. This is considered an important marker of exercise intensity at which the transition from aerobic to anaerobic metabolism occurs. The LT value was determined through identification of the upward inflection point on the blood lactate concentration versus running speed curve. The animals were allowed to rest for at least 30 min in individual cages, with free access to water. After this period, the rats were submitted to an initial warm-up period of 10 min at low speed (3 m/min) to remove the excess blood lactate accumulated during the manipulation preceding the test (Langfort et al. [1996](#page-12-0)). After a passive recovery during 3 min they were submitted to the IT, starting at the speed of 6 m/min, with an increase of 3 m/min at the end of every 3-min stage

(Takahashi et al. [2012](#page-13-0)). The IT finished when the animal reached exhaustion. Blood samples were collected from the tail vein between each stage. Lactate concentration was determined by an enzymatic–amperometric method, using a Lactate Scout analyzer (SensLab GmbH, Germany). Blood lactate concentration at different steps was plotted as a function of the corresponding running speed. Results are given as mmol/L.

### Measurement of amino acid levels

The plasma free AA levels change with exercise, in particular branched chain-AA (BCAA) and aromatic AA (AAAr). BCAAs, which include leucine (Leu), isoleucine (Ile) and valine (Val), are readily metabolized in the muscle. AAAr, including phenylalanine (Phe), tryptophan (Trp) and tyrosine, are degraded into the liver. AA concentrations were assayed using a high-performance liquid chromatography (HPLC) procedure as described by Donati et al. [\(2009](#page-11-0)). AA separation was carried out on a  $4.6 \times 250$  mm Bio-Sil ODS-5S column (particle size, 5 mm) in a Beckman HPLC system (equipped with 32 Karat software). AAs were determined by measuring the fluorescence of dansylated derivative with a Jasco spectrofluorometer (340 nm excitation, 525 emission). Norvaline was added as an internal standard to all samples. Levels of BCAA and AAAr are given as mmol/L.

Soleus and tibialis anterior protein carbonyl content

The measurement of protein carbonyl (PC) content has been used as a sensitive assay for oxidative damage to proteins in animal tissue. Muscles were weighed and diluted 20% w/v in potassium phosphate buffer (pH 6.7) with EDTA, and homogenized with an AEG SB2E-680, Germany. The introduction of PC groups into proteins by oxidative mechanisms was assayed by reaction of PC groups with primary amines to form semi-stable Schiff bases through reaction with 2,4 dinitrophenylhydrazine (DNPH), as described by Levine et al. [\(1990](#page-12-0)). PC content was calculated using the DNPH molar extinction coefficient (22,000/M/ cm). Results are given as nmol/mg protein.

### Statistical analysis

The analysis of variance (ANOVA) test was used to evaluate differences among multiple conditions. If positive, the Tukey test was used to test for their statistical significance. Student's  $t$  test was used to evaluate differences between two conditions. Values of  $P < 0.05$  were considered to be statistically significant.

# Results

# Rats' characteristics

Data related to body weight and food intake of the rats during the experiment are presented in Table 1. EOD treatment significantly lowered body weight and food intake. In addition to a significant overall age and EOD effect, there was also an age and EOD interaction both in body weight and in food intake. These data were not modified by exercise training programme. Survival percentage of EOD animals at the end of study was 90% and was not changed by training, whereas that of ALs rats was 50% and that of ALe rats was 70%.

# Body composition

Body composition changes are shown in Fig. [1.](#page-5-0) EOD rats kept the body fat content lower compared to AL rats up the end of protocol. No significant differences in body composition between sedentary and treadmilltrained groups were observed, although EODe rats showed an increase in fat free mass respect to EODs rats.

# Grasping test

Data obtained from the grasping test are presented in Fig. [2.](#page-5-0) The grip strength normalized to the animal body weight was similar for the AL rats during the experiment period. EOD rats had values higher respect to AL rats and the difference increased significantly at 24 months of age ( $P < 0.01$ ).

# Plasma analysis

# TAC

At 19 months of age, before exercise period, plasma TAC concentration in the food-restricted rats was

	ALs	ALe	<b>EODs</b>	EODe
Body weight $(g)^{a,b,c}$				
6 months	$529 \pm 7^{\rm{dw}}$		$451 \pm 5^{dy}$	
12 months	$627 \pm 10^{ew}$		$497 \pm 6^{e}$	
19 months	$673 \pm 10^{\rm fw}$	$673 \pm 10^{dw}$	$507 \pm 8$ <sup>ey</sup>	$507 \pm 8$ <sup>dy</sup>
24 months	$633 \pm 16^{\text{efw}}$	$630 \pm 14^{\rm{dw}}$	$508 \pm 7^{\rm ey}$	$485 \pm 8$ <sup>dy</sup>
Food intake $(g/day)^{a,b,c}$				
6 months	$21.6 \pm 0.3$ <sup>dw</sup>		$16.8 \pm 0.1$ <sup>dy</sup>	
12 months	$23.6 \pm 0.2^{\text{ew}}$		$17.6 \pm 0.2$ <sup>dy</sup>	
19 months	$24.2 \pm 0.7^{\rm ew}$	$24.2 \pm 0.7^{\text{dw}}$	$16.5 \pm 0.2$ <sup>dy</sup>	$16.5 \pm 0.2$ <sup>dy</sup>
24 months	$22.3 \pm 0.5^{\rm dew}$	$22.0 \pm 0.5^{\text{dw}}$	$16.5 \pm 0.4$ <sup>dy</sup>	$16.1 \pm 0.4^{\rm dy}$

Table 1 Body weight and food consumption in Sprague–Dawley rats submitted to different diet regimens and exercise

Values represent the mean  $\pm$  SEM

ALs ad libitum sedentary group, ALe ad libitum exercised group, EODs diet restricted sedentary group, EODe diet restricted exercised group

<sup>a</sup>Significant age effect ( $P < 0.01$ )

 $b$ Significant diet effect ( $P < 0.01$ )

<sup>c</sup>Significant age by diet effect ( $P < 0.01$ )

 $\text{det}$  Means in the same column across age groups with different superscripts are significantly different ( $P < 0.05$ )

<sup>wy</sup>Means in the same row across diet groups with different superscripts are significantly different ( $P < 0.05$ )

<span id="page-5-0"></span>Fig. 1 Changes in body composition in Sprague– Dawley rats submitted to different diet regimens and exercise. Results are given as percent changes in body weight. Results represent the mean of at least five cases. FAT fat mass, FFM fat free mass, ALs ad libitum sedentary group, ALe ad libitum exercised group, EODs diet restricted sedentary group, EODe diet restricted exercised group



Fig. 2 Grasping force in Sprague–Dawley rats submitted to different diet regimens and exercise. Results represent the mean  $\pm$  SEM of at least five cases. Two-way ANOVA statistical analysis (age  $\times$  diet) in sedentary rats—age main effect: N.S; diet main effect: N.S.; age by diet interaction: N.S.

**Quotient grasping strenght/body weight**

Quotient grasping strenght/body weight

significantly less than in the fed ad libitum (AL:  $0.46 \pm 0.04$  mmol Trolox equivalent/L; EOD:  $0.33 \pm 0.02$  mmol Trolox equivalent/L;  $P < 0.05$ ).

Two-way ANOVA statistical analysis (diet  $\times$  exercise)—\*\*diet main effect:  $(P < 0.01)$ ; exercise main effect: N.S. diet by exercise interaction: N.S. ALs ad libitum sedentary group, ALe ad libitum exercised group, EODs diet restricted sedentary group, EODe diet restricted exercised group

Figure [3](#page-6-0) shows that the effect of diet treatment was significant until 24 months of age; no significant agerelated change was observed, although there was a

<span id="page-6-0"></span>

Fig. 3 Effects of diet regimens and exercise on total antioxidant capacity (TAC) plasma concentration in Sprague–Dawley rats. Results represent the mean  $\pm$  SEM of at least five cases. Two-way ANOVA statistical analysis (age  $\times$  diet) in sedentary rats—age main effect: N.S; \*diet main effect:  $(P < 0.05)$ ; age by diet interaction: N.S. Two-way ANOVA statistical analysis

trend toward decreasing in AL rats. Exercise training programme lowered significantly plasma TAC levels in AL and EOD rats ( $P \lt 0.01$ ), and no significant effect of diet treatment was detected.

# LT

Data obtained on evaluation of blood LT are presented in Fig. 4. The lactate concentration at rest was similar in all groups. The velocity at LT was 9 m/min for EODs and ALs groups, 12 m/min for ALe group, and 18 m/min for EODe group. ALs group had reached exhaustion at 15 m/min, whereas ALe and EODs groups reached exhaustion at 18 m/min and only rats in EODe group run until 21 m/min. At the end of the IT test the blood lactate concentrations were lower in the trained groups than in the sedentary ones  $(P < 0.05)$ .

AAs

AA levels are showed in Fig. [5.](#page-7-0) BCAA levels at 19 months of age were significantly higher in EOD than AL rats ( $P < 0.01$ , only Ile was not statistically

(diet  $\times$  exercise)—diet main effect: N.S.; <sup>##</sup>exercise main effect:  $(P < 0.01)$ ; diet by exercise interaction: N.S. ALs ad libitum sedentary group, ALe ad libitum exercised group, EODs diet restricted sedentary group, EODe diet restricted exercised group



Fig. 4 Determination of plasma lactate threshold in Sprague– Dawley rats submitted to different diet regimens and exercise. Results represent the mean  $\pm$  SEM of at least five cases (error bars are the same size or smaller than the symbols). Two-way ANOVA statistical analysis (diet  $\times$  exercise)—diet main effect: N.S.;  $*$ exercise main effect: ( $P < 0.05$ ); diet by exercise interaction: N.S. ALs ad libitum sedentary group, ALe ad libitum exercised group, EODs diet restricted sedentary group, EODe diet restricted exercised group

significant). At 24 months of age, a significant decrease in BCAA values was observed in AL and

<span id="page-7-0"></span>

Fig. 5 Effects of diet regimens and exercise on branched chain amino acids (BCAA), aromatic amino acids (AAAr) and BCAA/AAAr ratio in plasma of Sprague–Dawley rats. Results represent the mean  $\pm$  SEM of at least five cases. Two-way ANOVA statistical analysis (age  $\times$  diet) in sedentary rats: BCAA Tot— $\degree$  age main effect: (P < 0.01); \*diet main effect:  $(P \lt 0.05)$ ; age by diet interaction:  $(P \lt 0.05)$ ; Post-ANOVA Tukey test ( $P < 0.05$ ): 19 months versus 24 months; AL versus EOD. Ile: age main effect:  $(P < 0.01)$ ; diet main effect: N.S.; age by diet interaction:  $(P < 0.05)$ ; Post-ANOVA Tukey test  $(P < 0.05)$ : 19 months versus 24 months. Leu: age main effect:  $(P < 0.01)$ ; diet main effect:  $(P < 0.01)$ ; age by diet interaction: N.S. Val: age main effect:  $(P < 0.01)$ ; diet main effect: N.S; age by diet interaction:  $(P < 0.01)$ ; Post-ANOVA Tukey test  $(P < 0.05)$ : 19 months versus 24 months AAAr Tot—age main effect: N.S.; diet main effect: N.S.; age by diet interaction:  $(P < 0.05)$ . Phe: age main effect:  $(P < 0.05)$ ; diet main effect:

N.S.; age by diet interaction:  $(P \lt 0.05)$ . Post-ANOVA Tukey test  $(P < 0.05)$ : 19 months versus 24 months BCAA/AAAr ratio— $\degree$ age main effect: (P < 0.01); diet main effect: N.S.; age by diet interaction: N.S Two-way ANOVA statistical analysis (diet  $\times$  exercise): BCAA Tot—diet main effect: N.S.; exercise main effect:  $(P < 0.05)$ ; diet by exercise interaction: N.S. Leu: diet main effect:  $(P < 0.05)$ ; exercise main effect:  $(P \lt 0.05)$ ; age by diet interaction: N.S AAAr Tot—diet main effect: N.S.;  $\sqrt[4]{\text{exercise}}$  main effect: ( $P < 0.05$ ); diet by exercise interaction: N.S. Phe: diet main effect: N.S.; exercise main effect:  $(P < 0.01)$ ; diet by exercise interaction: N.S. *BCAA*/ AAAr ratio—\*\*diet main effect:  $(P < 0.01)$ ; exercise main effect: N.S.; diet by exercise interaction: N.S. Ile isoleucine, Leu leucine, Val valine, Phe phenylalanine, Trp tryptophan, Tyr tyrosine, ALs ad libitum sedentary group, ALe ad libitum exercised group, EODs diet restricted sedentary group, EODe diet restricted exercised group

EOD groups  $(P < 0.01)$ , and the effect of diet treatment was significant only in Leu levels  $(P < 0.01)$ . Exercise training programme lowered significantly BCAA levels in AL and EOD rats  $(P < 0.05)$ , and the effect of diet treatment was significant only in Leu levels ( $P \leq 0.05$ ).

As far as AAAr are concerned, there were no significant differences in the Trp and Tyr plasma levels in all experimental groups, while Phe values showed a significant decrease with aging  $(P < 0.05)$ and after exercise ( $P < 0.01$ ). A significant decrease in BCAA/AAAr ratio was observed between 19 and 24 months of age in ALs and EODs rats ( $P < 0.01$ ). After exercise training programme the ratio was significantly higher in EOD than AL rats ( $P < 0.01$ ).

### PC groups

The effects of exercise training programme on biomarker of muscle protein oxidation are presented in Fig. 6. PC groups were affected differentially by exercise and diet treatment, depending on muscle type. Exercise did not affect soleus PC levels either in AL or EOD rats, while difference was observed between diet groups: PC content in the AL groups was significantly lower than that in the EOD groups  $(P < 0.01)$ . Exercise induced a significant increase of PC groups in tibialis anterior muscle ( $P < 0.05$ ); the effect of diet treatment was not significant, although a more pronounced increase was observed in the AL group.

#### **Discussion**

In the present study, the effects on physiological and metabolic parameters of late-onset exercise training were investigated in rats with the same ''chronological age'' but different ''biological age'' as a result of different lifelong dietary regimens: the positive effects of DR on longevity and on health span are well known (Liang et al. [2018\)](#page-12-0), but the effects of exercise in late age have not been yet studied in detail.

### Physiological parameters

The pattern of changes over time in body weight, food intake and survival percentage are comparable to patters shown in our previous studies on DR effects (Cavallini et al. [2002;](#page-11-0) Bonelli et al. [2008](#page-11-0)). Exercise



Fig. 6 Effects of diet regimens and exercise on soleus and tibialis anterior protein carbonyl content in Sprague–Dawley rats. Results represent the mean  $\pm$  SEM of at least five cases. Two-way ANOVA statistical analysis (diet  $\times$  exercise): Soleus—\*\*diet main effect:  $(P < 0.01)$ ; exercise main effect: N.S.; diet by exercise interaction: N.S. tibialis anterior—diet main effect: N.S.;  $*$ exercise main effect: ( $P < 0.05$ ); diet by exercise interaction: N.S. Sed sedentary groups, Exe exercised groups, AL ad libitum rats, EOD restricted rats

training programme did not cause significant changes, only the survival percentage in AL rats showed an increase: given that our findings is based on a limited number of animals (10 rats for experimental group), the result from such analyses should thus be treated with caution. However, these data support the idea that a regular exercise may beneficial affect and delay biological aging (Radak et al. [2005](#page-12-0)), even when initiated in old animals (Cobley et al. [2015](#page-11-0)). The body composition was measured by BIA and was comparable to patterns shown in other studies on the same rat strain (Skalicky et al. [2001\)](#page-13-0): after 19 months of age the fat content tended to stay stable, exercise caused a

slight increase in free fat mass only EOD rats, probably DR preserves lean body mass from agerelated modifications as observed by McKiernan et al. [\(2012](#page-12-0)) in aged rhesus monkeys. Therefore, muscle fibers from EOD rats might be better poised to endure and adapt to changes like exercise training programme. Interestingly, data obtained with grasping test further support the protective role of DR on skeletal muscle mass: indeed, EOD rats showed a higher grip strength respect to AL rats until 24 months, and the training improved their grip strength. The age-related loss of muscle strength has been defined as dynapenia and was related to deficits in muscle quality and neuromuscular control (Manini and Clark [2012](#page-12-0)). It is known that the anti-aging effect of DR involved the activation of autophagy, a required function in cell housekeeping during fasting, which can remove damaged macromolecules, organelles and membranes selectively, acting as an alternative source of energy and participating in cell quality control (Bergamini et al. [2007](#page-11-0); Hubbard et al. [2012](#page-12-0)). A recent paper suggests that autophagy is required for exercise training-induced skeletal muscle adaptation and improvement of physical performance, and this cell function is very active and more intense when exercise is performed in fasted state (Martin-Rincon et al. [2018\)](#page-12-0).

### Plasma metabolic parameters

TAC is a sensitive and reliable marker for detecting plasma changes in vivo oxidative stress that may not be detectable through the measurement of a single, specific antioxidant (Erel [2004](#page-11-0)), and is used to evaluate a number of physiological conditions in humans and animals (Ghiselli et al. [2000](#page-11-0)). An interesting finding in this study was that plasma TAC concentrations in EOD rats were less than in AL rats. On the basis of such data, it seems reasonable to suggest that DR-induced decrease in reactive oxygen species (ROS) production may result in a lessening of the requirement for TAC. It has been reported that exercise decreased plasma TAC in rats: our results are in line with these findings (Ficicilar et al. [2003](#page-11-0)). A growing number of reports indicate that exerciseinduced ROS production is required to promote exercise training response in skeletal muscle and contributes to exercise-induced skeletal muscle adaptation (Davies et al. [1982](#page-11-0); Gomez-Cabrera et al. [2008](#page-12-0)).

Blood LT, defined as the point at which blood lactate concentration increases exponentially with increasing exercise intensity, has shown to be a useful tool in the exercise prescription (Billat et al. [2003](#page-11-0)). Many researches on exercise physiology have been conducted with laboratory animals and the ''aerobic/ anaerobic'' transition has been used to ascertain endurance capability and measure adaptations to training (Gobatto et al. [2001](#page-12-0); Billat et al. [2004\)](#page-11-0). Our incremental training was found to be efficient in improving physical fitness of the rats in 8 weeks. The late lactate increase in trained animals suggests improvement of the aerobic metabolism (Sjödin and Jacobs [1981](#page-13-0)), and the increase in the time of exhaustion indicates improvement in the cardiac performance, through increased cardiac output (Li et al. [2018\)](#page-12-0). Mammalian skeletal muscle fibers are subject to significant changes during postnatal development and aging (Schiaffino and Reggiani [2011\)](#page-13-0) and lactate levels are known to be influenced by the muscle fiber composition (Kitada et al. [2015](#page-12-0)). During aging, rat skeletal muscles undergo a type 2B to 2X switching in fast muscles and a type 2A to type 1 switching in slow muscles (Larsson et al. [1995\)](#page-12-0). Previous studies have reported a strong relationship between the number of slow muscle fibers and LT value (Ivy et al. [1980\)](#page-12-0). As expected, EOD rats shown better performance than AL rats: probably the fiber type transitions increase the amount of oxidative type I fibers which are more resistant to fasting than type II glycolytic fibers (Wang and Pessin [2013](#page-13-0)).

Among plasma free AAs, BCAA (Ile, Leu and Val) are key regulators of protein synthesis. Unlike other AAs, BCAA are not metabolized in the liver, and therefore after the ingestion they are almost immediately put into circulation and made available to the body (Dato et al. [2019\)](#page-11-0). BCAA catabolism is mainly located in skeletal muscle and the brain, but also adipose tissue can metabolize substantial amounts of BCAA (Herman et al. [2010\)](#page-12-0). This factor may be responsible for different BCAA plasma levels in AL and EOD rats at the beginning of the experiment: AL rats showed more FFA than EOD, after 19 months of age the fat content tended to remain stable. Age and exercise training lowered significantly BCAA in AL and EOD rats. It is interesting to note that a diet significant effect was observed only on Leu levels: probably the different meal distribution between groups might affect the Leu plasma levels, as suggested by Norton et al. ([2017\)](#page-12-0). In addition, this result highlights that the protective role of DR on skeletal muscle mass might be associated with an effect on Leu levels: Leu is the main stimulator for protein synthesis in the skeletal muscle and improves whole body glucose metabolism, with action on glucose muscle uptake, body weight and food intake (Valerio et al. [2011\)](#page-13-0). Furthermore, the decrease in the BCAA plasma concentrations may be relate to fatigue, especially in old animals. Indeed, no significant changes was observed in Trp levels: Trp is a precursor of serotonin, and some studies have shown that excessive serotonin induced fatigue (Cordeiro et al. [2017\)](#page-11-0). Since the transporter for Trp and BCAA is the same (Fernstrom [2005\)](#page-11-0), when BCAA levels decrease, a larger amount of Trp can entry into the brain and cause fatigue. In this perspective, also BCAA/AAAr ratio showed an age-related decrease: the ratio was significantly higher in EOD rats than AL rats. Unlike BCAA, AAAr undergo liver metabolism process and BCAA/AAAr ratio is used as an indicator of liver function (Holecek et al. [1996](#page-12-0)): the liver of EOD rats might be affected by ''anticipatory activity'', when animals show increased locomotor activity 2–3 h before food access, which depend on a food-entrain-able oscillator (Díaz-Muñoz et al. [2000\)](#page-11-0). The authors suggest that hepatic metabolism in DR rats is modulated with a different pattern from AL rats, and during "anticipatory activity" the liver of food-restricted animals optimizes the processing of nutrients to daily feeding with an anticipatory function in the control of energy balance. Furthermore, DR may enhance liver autophagy during the longer time period of fasting and remove damaged macromolecules, organelles and membranes selectively, acting as an alternative source of energy (Donati et al. [2001\)](#page-11-0).

### PC groups

PC groups are well known as a useful and reliable marker for assessing oxidative stress in skeletal muscle (Çakatay et al. [2003](#page-11-0)). Our findings show that this marker was affected differentially by exercise and diet treatment, depending on muscle type. Unexpectedly, levels of PC were higher EOD rats, especially in soleus muscle. DR animals are more active than AL animals at all ages, as reported by several studies (Duffy et al. [1989;](#page-11-0) Yamada et al. [2013\)](#page-13-0). Increased protein oxidation might be associated with an elevated metabolic rate in muscle tissue involved in spontaneous activity. DR increases the turnover of cell components and disposal of damaged protein or organelles by autophagic and proteasomal degradation (Bonelli et al. [2008](#page-11-0); Hubbard et al. [2012](#page-12-0)): therefore, the accumulation of damaged protein should be reduced. The age-related muscle fiber type transitions and metabolic shifts in aging muscle can offer an explanation for the increase of PC groups (Larsson et al. [1995](#page-12-0); Schiaffino and Reggiani [2011\)](#page-13-0), especially in slow-twitch muscles, rich in myoglobin and oxidative enzymes. Furthermore, PC groups may be considered intermediate products of oxidation, since further oxidation and cross-linking results in the formation of fluorescent age pigments (Sitte et al. [2000\)](#page-13-0). In the AL animals, where autophagic and proteasomal degradation are impaired, oxidized proteins might remain more time within cells and be subjected to more modifications and cross-linking, lowering free PC groups to be detected by the assay. However, exercise does not seem to change a pattern already defined by age and diet treatment, although in EOD soleus muscle was observed a decrease of PC groups, probably training might further improve the activity of proteasomal complex (Radak et al. [2019](#page-12-0)). The increase of PC levels in AL and EOD tibialis anterior muscle may highlight the major susceptibility of type II fibers to aging, both in natural aging than accelerated-mimetic aging models (Wang and Pessin [2013;](#page-13-0) Yanar et al. [2019](#page-13-0)).

#### Conclusion

A considerable amount of knowledge regarding the relationship between exercise and aging is derived from animal research, particularly rodents. In this regard, our results show that late-onset exercise training has the potential to improve performance and metabolic parameters in rats with the same ''chronological age'' but different ''biological age''. Furthermore, our results indicate that physical exercise does not have negative effect on the majority of the testing parameters, does not increase the difference between animals with different ''biological age'', and might be useful to separate the influence of exercise from those that occur solely due to aging. Interestingly, a recent study shows that rat responses to exercise adequately reflect human ones in blood <span id="page-11-0"></span>parameters linked to various organs, tissues, functions, and diseases (Goutianos et al. [2015](#page-12-0)). Based on such data, our results might represent a spur for future studies, using rat model to understand how exercise training, initiated in late middle age, may improve physical functions, and consequently quality of life of the elderly population. Many critical questions still remain regarding the relationship of aging and exercise, but our results highlight the relevance of a personalised and selected exercise protocol, since the responsiveness to exercise may depend on the individual's "biological age".

Acknowledgements We thank Silvia Giacomelli for her assistance in exercising the animals.

Author contributions All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by SC, MD and GC. The first draft of the manuscript was written by GC and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

#### Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

### References

- Alessio HM, Hagerman AE, Fulkerson BK, Ambrose J, Rice RE, Wiley RL (2000) Generation of reactive oxygen species after exhaustive aerobic and isometric exercise. Med Sci Sports Exerc 32:1576–1581
- Ben J, Soares FM, Cechetti F, Vuaden FC, Bonan CD, Netto CA, Wyse AT (2009) Exercise effects on activities of  $Na(+)$ ,  $K(+)$ -ATPase, acetylcholinesterase and adenine nucleotides hydrolysis in ovariectomized rats. Brain Res 1302:248–255
- Ben J, Soares FM, Scherer EB, Cechetti F, Netto CA, Wyse AT (2010) Running exercise effects on spatial and avoidance tasks in ovariectomized rats. Neurobiol Learn Mem 94:312–317
- Bergamini E, Cavallini G, Donati A, Gori Z (2007) The role of autophagy in ageing: its essential part in the anti-ageing mechanism of caloric restriction. Ann N Y Acad Sci 1114:69–78
- Billat VL, Sirvent P, Py G, Koralsztein JP, Mercier J (2003) The concept of maximal lactate steady state: a bridge between biochemistry, physiology and sport science. Sports Med 33:407–426
- Billat VL, Mouisel E, Roblot N, Melki J (2004) Inter- and intrastrain variation in mouse critical running speed. J Appl Physiol 98:1258–1263
- Bonelli MA, Desenzani S, Cavallini G, Donati A, Romani AA, Bergamini E, Borghetti AF (2008) Low-level caloric

restriction rescues proteasome activity and Hsc70 level in liver of aged rats. Biogerontology 9:1–10

- Çakatay U, Telci A, Kayali R, Tekeli F, Akçay T, Sivas A (2003) Relation of aging with oxidative protein damage parameters in the rat skeletal muscle. Clin Biochem 36:51–55
- Carvalho JF, Masuda MO, Pompeu FA (2005) Method for diagnosis and control of aerobic training in rats based on lactate threshold. Comp Biochem Physiol Part A 140:409–413
- Cavallini G, Donati A, Gori Z, Parentini I, Bergamini E (2002) Low level dietary restriction retards age-related dolichol accumulation. Aging Clin Exp Res 14:152–154
- Chow LS, Greenlund LJ, Asmann YW, Short KR, McCrady SK, Levine JA, Nair KS (2007) Impact of endurance training on murine spontaneous activity, muscle mitochondrial DNA abundance, gene transcripts, and function. J Appl Physiol 102:1078–1089
- Cobley JN, Moult PR, Burniston JG, Morton JP, Close GL (2015) Exercise improves mitochondrial and redox-regulated stress responses in the elderly: better late than never! Biogerontology 16:249–264
- Cordeiro LMS, Rabelo PCR, Moraes MM, Teixeira-Coelho F, Coimbra CC, Wanner SP, Soares DD (2017) Physical exercise-induced fatigue: the role of serotonergic and dopaminergic systems. Braz J Med Biol Res 50:e6432
- Dato S, Hoxha E, Crocco P, Iannone F, Passarino G, Rose G (2019) Amino acids and amino acid sensing: implication for aging and diseases. Biogerontology 20:17–31
- Davies KJ, Quintanilha AT, Brooks GA, Packer L (1982) Free radicals and tissue damage produced by exercise. Biochem Biophys Res Commun 107:1198–1205
- Díaz-Muñoz M, Vázquez-Martínez O, Aguilar-Roblero R, Escobar C (2000) Anticipatory changes in liver metabolism and entrainment of insulin, glucagon, and corticosterone in food-restricted rats. Am J Physiol Regul Integr Comp Physiol 279:R2048–2056
- Donati A, Cavallini G, Paradiso C, Vittorini S, Pollera M, Gori Z, Bergamini E (2001) Age-related changes in the autophagic proteolysis of rat isolated liver cells: effects of antiaging dietary restrictions. J Gerontol A Biol Sci Med Sci 56:B375–383
- Donati A, Cavallini G, Bergamini E (2009) Methods for inducing and monitoring liver autophagy relative to aging and antiaging caloric restriction in rats. Methods Enzymol 452:441–455
- Duffy PH, Feuers RJ, Leakey JA, Nakamura K, Turturro A, Hart RW (1989) Effect of chronic caloric restriction on physiological variables related to energy metabolism in the male Fischer 344 rat. Mech Ageing Dev 48:117–133
- Erel O (2004) A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. Clin Biochem 37:277–285
- Fernstrom JD (2005) Branched-chain amino acids and brain function. J Nutr 135:1539–1546
- Ficicilar H, Zergeroglu AM, Tekin D, Ersoz G (2003) The effects of acute exercise on plasma antioxidant status and platelet response. Thromb Res 111:267–271
- Ghiselli A, Serafini M, Natella F, Scaccini C (2000) Total antioxidant capacity as a tool to assess redox status: critical view and experimental data. Free Radic Biol Med 29:1106–1114
- <span id="page-12-0"></span>Gobatto CA, de Mello MA, Sibuya CY, de Azevedo JR, dos Santos LA, Kokubun E (2001) Maximal lactate steady state in rats submitted to swimming exercise. Comp Biochem Physiol A 130:21–27
- Gomez-Cabrera MC, Domenech E, Romagnoli M, Arduini A, Borras C, Pallardo FV, Sastre J, Vina J (2008) Oral administration of vitamin C decreases muscle mitochondrial biogenesis and hampers training-induced adaptations in endurance performance. Am J Clin Nutr 87:142–149
- Goto S, Radák Z (2009) Hormetic effects of reactive oxygen species by exercise: a view from animal studies for successful aging in human. Dose Response 8:68–72
- Goutianos G, Tzioura A, Kyparos A, Paschalis V, Margaritelis NV, Veskoukis AS, Zafeiridis A, Dipla K, Nikolaidis MG, Vrabas IS (2015) The rat adequately reflects human responses to exercise in blood biochemical profile: a comparative study. Physiol Rep 3:e12293
- Heilbronn LK, de Jonge L, Frisard MI, DeLany JP, Larson-Meyer DE, Rood J, Nguyen T, Martin CK, Volaufova J, Most MM, Greenway FL, Smith SR, Deutsch WA, Williamson DA, Ravussin E, Pennington CALERIE Team (2006) Effect of 6-month calorie restriction on biomarkers of longevity, metabolic adaptation, and oxidative stress in overweight individuals: a randomized controlled trial. JAMA 295:1539–1548
- Herman MA, She P, Peroni OD, Lynch CJ, Kahn BB (2010) Adipose tissue branched chain amino acid (BCAA) metabolism modulates circulating BCAA levels. J Biol Chem 285:11348–11356
- Herndon LA, Schmeissner PJ, Dudaronek JM, Brown PA, Listner KM, Sakano Y, Paupard MC, Hall DH, Driscoll M (2002) Stochastic and genetic factors influence tissuespecific decline in ageing C. elegans. Nature 419:808–814
- Holecek M, Mraz J, Tilser I (1996) Plasma amino acids in four models of experimental liver injury in rats. Amino Acids 10:229–241
- Holloszy JO, Smith EK, Vining M, Adams S (1985) Effect of voluntary exercise on longevity of rats. J Appl Physiol 59:826–831
- Hubbard VM, Valdor R, Macian F, Cuervo AM (2012) Selective autophagy in the maintainance of cellular homeostasis in aging organism. Biogerontology 13:21–35
- Ivy JL, Withers RT, Van Handel PJ, Elger DH, Costill DL (1980) Muscle respiratory capacity and fiber type as determinants of the lactate threshold. J Appl Physiol Respir Environ Exerc Physiol 48:523–527
- Kennedy BK, Steffen KK, Kaeberlein M (2007) Ruminations on dietary restriction and aging. Cell Mol Life Sci 64:1323–1328
- Kennedy BK, Berger SL, Brunet A, Campisi J, Cuervo AM, Epel ES, Franceschi C, Lithgow GJ, Morimoto RI, Pessin JE, Rando TA, Richardson A, Schadt EE, Wyss-Coray T, Sierra F (2014) Geroscience: linking aging to chronic disease. Cell 159:709–713
- Kitada T, Machida S, Naito H (2015) Influence of muscle fibre composition on muscle oxygenation during maximal running. BMJ Open Sport Exerc Med 1:e000062
- Langfort J, Zarzeczny R, Pilis W, Kaciuba-Uściłko H, Nazar K, Porta S (1996) Effect of sustained hyperadrenalinemia on exercise performance and lactate threshold in rats. Comp Biochem Physiol Part A 114:51–55
- Larsson L, Müller U, Li X, Schiaffino S (1995) Thyroid hormone regulation of myosin heavy chain isoform composition in young and old rats, with special reference to IIX myosin. Acta Physiol Scand 153:109–116
- Lefevre M, Redman LM, Heilbronn LK, Smith JV, Martin CK, Rood JC, Greenway FL, Williamson DA, Smith SR, Ravussin E, Pennington CALERIE team (2009) Caloric restriction alone and with exercise improves CVD risk in healthy non-obese individuals. Atherosclerosis 203:206–213
- Levine RL, Garland D, Oliver CN, Amici A, Climent I, Lenz AG, Ahn BW, Shaltiel S, Stadtman ER (1990) Determination of carbonyl content in oxidatively modified proteins. Methods Enzymol 186:464–478
- Li FH, Li T, Su YM, Ai JY, Duan R, Liu TC (2018) Cardiac basal autophagic activity and increased exercise capacity. J Physiol Sci 68:729–742
- Liang Y, Liu C, Lu M, Dong Q, Wang Z, Wang Z, Xiong W, Zhang N, Zhou J, Liu Q, Wang X, Wang Z (2018) Calorie restriction is the most reasonable anti-ageing intervention: a meta-analysis of survival curves. Sci Rep 8:5779
- Manini TM, Clark BC (2012) Dynapenia and aging: an update. J Gerontol A Biol Sci Med Sci 67:28–40
- Martin-Rincon M, Morales-Alamo D, Calbet JAL (2018) Exercise-mediated modulation of autophagy in skeletal muscle. Scand J Med Sci Sports 28:772–781
- Masoro EJ (1980) Mortality and growth characteristics of rat strains commonly used in aging research. Exp Aging Res 6:219–233
- Mattison JA, Colman RJ, Beasley TM, Allison DB, Kemnitz JW, Roth GS, Ingram DK, Weindruch R, de Cabo R, Anderson RM (2017) Caloric restriction improves health and survival of rhesus monkeys. Nat Commun 8:14063
- McKiernan SH, Colman RJ, Aiken E, Evans TD, Beasley TM, Aiken JM, Weindruch R, Anderson RM (2012) Cellular adaptation contributes to calorie restriction-induced preservation of skeletal muscle in aged rhesus monkeys. Exp Gerontol 47:229–236
- Norton LE, Wilson GJ, Moulton CJ, Layman DK (2017) Meal distribution of dietary protein and leucine influences long term muscle mass and body composition in adult rats. J Nutr 147:195–201
- Pilis W, Zarzeczny R, Langfort J, Kaciuba-Uściłko H, Nazar K, Wojtyna J (1993) Anaerobic threshold in rats. Comp Biochem Physiol Comp Physiol 106:285–289
- Powers SK, Radak Z, Ji LL (2016) Exercise-induced oxidative stress: past, present and future. J Physiol 594:5081–5092
- Radak Z, Tolvaj D, Ogonovszky H, Toldy A, Taylor AW (2004) Exercise and cancer. In: Radak Z (ed) Exercise and diseases. Meyer Meyer Sport, Oxford, pp 168–190
- Radak Z, Chung HY, Goto S (2005) Exercise and hormesis: oxidative stress-related adaptation for successful aging. Biogerontology 6:71–75
- Radak Z, Torma F, Berkes I, Goto S, Mimura T, Posa A, Balogh L, Boldogh I, Suzuki K, Higuchi M, Koltai E (2019) Exercise effects on physiological function during aging. Free Radic Biol Med 132:33–41
- Rattan SI (2015) Biology of ageing: principles, challenges and perspectives. Rom J Morphol Embryol 56:1251–1253
- Sahlin K, Shabalina IG, Mattsson CM, Bakkman L, Fernström M, Rozhdestvenskaya Z, Enqvist JK, Nedergaard J,

<span id="page-13-0"></span>Ekblom B, Tonkonogi M (2010) Ultraendurance exercise increases the production of reactive oxygen species in isolated mitochondria from human skeletal muscle. J Appl Physiol 108:780–787

- Schiaffino S, Reggiani C (2011) Fiber types in mammalian skeletal muscles. Physiol Rev 91:1447–1531
- Sitte N, Merker K, Von Zglinicki T, Davies KJ, Grune T (2000) Protein oxidation and degradation during cellular senescence of human BJ fibroblasts: part II-aging of nondividing cells. FASEB J 14:2503–2510
- Sjödin B, Jacobs I (1981) Onset of blood lactate accumulation and marathon running performance. Int J Sports Med 2:23–26
- Skalicky M, Narath E, Viidik A (2001) Housing conditions influence the survival and body composition of ageing rats. Exp Gerontol 36:159–170
- Smith JP, Hicks PS, Ortiz LR, Martinez MJ, Mandler RN (1995) Quantitative measurement of muscle strength in the mouse. J Neurosci Methods 62:15–19
- Spiering BA, Kraemer WJ, Anderson JM, Armstrong LE, Nindl BC, Volek JS, Maresh CM (2008) Resistance exercise biology: manipulation of resistance exercise programme variables determines the responses of cellular and molecular signalling pathways. Sports Med 38:527–540
- Takahashi H, Himi N, Kuniyasu K, Koga T (2012) Changes in the lactate threshold during treadmill exercise after microsphere-induced infarction in rats. J Stroke Cerebrovasc Dis 21:647–651
- Valerio A, D'Antona G, Nisoli E (2011) Branched-chain amino acids, mitochondrial biogenesis, and healthspan: an evolutionary perspective. Aging (Albany NY) 3:464–478
- Wang Y, Pessin JE (2013) Mechanisms for fiber-type specificity of skeletal muscle atrophy. Curr Opin Clin Nutr Metab Care 16:243–250
- Westerterp KR (2013) Physical activity and physical activity induced energy expenditure in humans: measurement, determinants, and effects. Front Physiol 4:90
- Westerterp KR (2015) Daily physical activity as determined by age, body mass and energy balance. Eur J Appl Physiol 115:177–1184
- Yamada Y, Colman RJ, Kemnitz JW, Baum ST, Anderson RM, Weindruch R, Schoeller DA (2013) Long-term calorie restriction decreases metabolic cost of movement and prevents decrease of physical activity during aging in rhesus monkeys. Exp Gerontol 48:1226–1235
- Yanar K, Simsek B, Atukeren P, Aydin S, Çakatay U (2019) Is d-galactose a useful agent for accelerated aging model of gastrocnemius and soleus muscle of sprague-dawley rats? Rejuvenation Res. 1:2. [https://doi.org/10.1089/rej.2019.](https://doi.org/10.1089/rej.2019.2185) [2185](https://doi.org/10.1089/rej.2019.2185)

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