The conversion of redox status of peritoneal macrophages during pathological progression of spontaneous inflammatory bowel disease in Janus family tyrosine kinase $3^{-/-}$ and IL-2 receptor $\gamma^{-/-}$ mice

Yukie Murata¹, Akira Yamashita², Takashi Saito³, Kazuo Sugamura⁴ and Junji Hamuro¹

¹Basic Research Laboratories, Ajinomoto Co. Inc., 1-1 Suzuki-cho, Kawasaki-ku, Kawasaki 210-0861, Japan

²Department of Anatomy, Hamamatsu Medical University, Hamamatsu 431-3192, Japan

³Division of Molecular Genetics, Chiba University School of Medicine, Chiba 263-8522, Japan

⁴Department of Immunology, Tohoku University School of Medicine, Sendai 980-8575, Japan

Keywords: glutathione, IL-2 receptor γ , inflammatory bowel disease, innate immunity, JAK3, reductive and oxidative macrophages

Abstract

The distinct thiol redox status in macrophages, either elevated or reduced intracellular content of glutathione (GSH), was confirmed during aging in IL-2 receptor (IL-2R) γ and Janus family tyrosine kinase (JAK)3 gene-disrupted mice. Oxidative macrophages (OMp) with reduced GSH dominated initially at a younger age in both mice. OMp-dominated JAK3 or IL-2R γ chain-deficient mice showed shortened life longevity compared with wild-type littermates. These mice elicited spontaneous onsets of inflammatory bowel disease (IBD)-like symptoms accompanied with the conversion of the redox status of macrophages to reductive phenotypes with elevated intracellular GSH. Conversion of OMp to the reductive phenotype by GSH monoethyl ester or by a β -(1–3)-glucan accelerated the disease onset, concomitant with the skewing from T_h2 to T_h1 responses. On the contrary, *N*,*N*-diacetyl cystine dimethylester, which is capable of inducing OMp, delayed the incidence of IBD-like symptoms and improved the survival rate. This implies that the conversion of OMp/T_h2 to reductive macrophages/T_h1 may be critical for the disease progression. The study of these mice may provide insight into the mechanisms underlying Crohn's disease and ulcerative colitis.

Introduction

The pathogenesis of inflammatory bowel disease (IBD), Crohn's disease (CD) and ulcerative colitis (UC) remains poorly understood (1). Initiation and progression of the intestinal inflammation in IBD may be a result of the dysregulated cellular immune responses of the intestinal mucosa to a constituent of endogenous luminal bacterial flora (2). Accumulating evidence indicates that activated CD4⁺ T lymphocytes are likely to play a central role in the pathogenesis of IBD (3). Several mouse models for IBD have already shown that alterations in cytokine propensities result from a failure of regulation by T_h cells (4–6). T_h1 cells produce pro-inflammatory cytokines such as IL-2 and IFN- γ , whereas T_h2 cells produce anti-inflammatory cytokines such as IL-4 and IL-10. In general, human CD is thought to be a T_h1 -like disease, whereas UC is thought to be a T_h2 -like disease (7–9). Moreover, crossbreeding IL-2 gene-disrupted mice with RAG2^{-/-} mice prevents colitis, whereas cross-breeding with JH-/- mice has no effect (10).

Macrophages play diverse and relevant roles in host defense against invasive and noxious insults (11,12). Recently, the important role of innate immune responses has been re-evaluated based on the findings of the essential roles

628 Pathogenesis of IBD and thiol redox status of macrophages



Fig. 1. Life longevity and onset of IBD-like diseases. (A) Life longevity of IL-2R^{-/Y} and JAK3^{-/-} male mice was followed using 48 and 49 mice respectively under normal breeding conditions. (B) IBD-like disease onset was evaluated by the occurrence of anal prolapse (n = 18 for IL-2R^{-/Y} and n = 20 for the other three strains). (C) Macroscopic inflammation score. At the age indicated, macroscopically visible inflammation was assessed using the scoring criteria described by Grisham *et al.* (51), with a maximum score of 11. A macroscopic score was obtained by summation of scores on severity of colonic adhesion to surrounding tissues, strictures, mucosal ulceration, hyperemia, mesentery bleeding and bowel wall thickness. The number shown in square brackets indicates the relative body weight (percent) compared with the wild-type mice of the same age. n.d., not done.

of IFN-y produced by macrophages and of the Toll-like receptor (TLR) family-MyD88-NF-κB system (13-15). Intestinal epithelial cell lines constitutively express several functional TLR, key regulators of the innate response system. Infiltrating macrophages and neutrophils are abundantly present in the inflamed gut of patients with IBD. The pivotal role of IL-12 produced by macrophages was demonstrated in chronic intestinal inflammation. There is a growing body of evidence that these cells expose the inflamed intestine to substantial oxidative stress by production of reactive oxygen species (ROS) and that ROS contribute to tissue injury in IBD (16–19). Intestinal inflammation induced by a ROS-generating agent could be reversed by the repletion of glutathione (GSH) (20). GSH is the most important intracellular anti-oxidant and is abundant in virtually all cells, playing significant roles in many biological processes (21). This has led to the proposal that IBD results from an imbalance between pro-oxidant and antioxidant mechanisms (22,23). Our group and others have demonstrated that the intracellular content of GSH in macro-

phages is critical for the secretion of IL-12 by regulating MAPK p38 activity (24-29). This suggests that GSH levels in macrophages play a crucial role in determining whether T_b1 or T_b2 cytokine responses predominate (27). Furthermore, the exposure of macrophages to IFN-y increased the GSH:oxidized GSH (GSSG), ratio whereas exposure of macrophages to IL-4 decreased the GSH:GSSG ratio (27,30). Thus, the ability to generate a T_h1 - or T_h2 -type response has turned out to depend not only on T cells, but also on the intracellular thiol redox status of macrophages (27). Macrophages with a reduced intracellular GSH content are referred to as oxidative macrophages (OMp) and those with an elevated amount are referred to as reductive macrophages (RMp) (24,26-28). The OMp showed elevated IL-6 and IL-10 production, and reduced NO and IL-12 production, while the RMp elicited a reciprocal response, i.e. elevated IL-12 and NO production, and reduced IL-6 and IL-10 production. The cytokine propensities of OMp or RMp were inter-converted to each other in the absence of cell proliferation and protein synthesis. Accumulating evidence suggests that intracellular GSH:GSSG balance regulates transcription factor NF-KB or AP-1 activity relevant for the gene activation of cytokines (31). The cytosolic complex cascade of phosphorylation by kinase and dephosphorylation by phosphatase is modulated by intracellular redox states (32). A paradigm has been proposed based on whether the presence of the functional heterogeneity of macrophages (33-36) is pro-inflammatory or anti-inflammatory. Our discussions treat RMp as pro-inflammatory and OMp as anti-inflammatory (27). This paper describes the pathogenesis of IBD in Janus family tyrosine kinase (JAK)3and IL-2 receptor (IL-2R)y-deficient mice and provides evidence that the $T_h 1/T_h 2$ balance of these mice is also regulated by the intracellular thiol redox status of macrophages. The stepwise skewing of the redox status in macrophages (oxidative to reductive) generated first T_b2 and then T_b1 dominancy, resulting in the periodical infiltration of inflammatory cells into intestinal mucosa, followed by tissue injury and anal prolapse along with disease progression. The successful prophylactic/therapeutic intervention with either cysteine or GSH derivatives is also discussed.

Methods

Mice and reagents

C57BL/6 mice were obtained from Charles River (Atsugi, Japan). JAK3 gene-disrupted (JAK3-/-) and IL-2Ry chain gene-disrupted (IL-2R $\gamma^{-/-}$) mice, both of the C57BL/6 background, were established as previously reported (37,38). Founders (F₀) of generated gene-disrupted mice were identified by PCR analysis of tail DNA. The primers used for genedisrupted mouse screening were as follows: forward 5'-ACCCAGGTACTCCATGCCCT-3' (JAK3). 5'-CTG-CTCAGAATGCCTCCAATTCC-3' (IL-2Ry); reverse 5'CCA-GACCAGCAGAGGGACTT-3' (JAK3), 5'CCTGCGTGCAATC-CATCTTGTTCAAT-3' (IL-2Ry); and reverse primers for an inserted neomycin gene 5'-CGACCACCAAGCGAAACATC-3' (JAK3), 5'-GATCCAGATTGCCAAGGTGAGTAG-3' (IL-2Rγ). The gene-disrupted mice were bred with either C57BL/6 or with homo- or hetero-knockout mice. GSH monoethylester IL2Ry +/Y

IL2R γ +/-





IL2Ry -/Y

IL2Rγ -/-

JAK3-/- (male)

Fig. 2. Histology of colonic inflammation. The age of each mouse was 13 weeks for IL-2R^{+//} and IL-2R^{-//}, 15 weeks for IL-2R^{+/-} and IL-2R^{-/-}, and 23 weeks for JAK3^{+/+} and JAK3^{-/-} (male).

(GSH-OEt) was purchased from Sigma (St Louis, MO). N,N-Diacetyl cystine dimethylester [(NACOMe)₂] was from Bachem (Bubendorf, Switzerland). Lipopolysaccharide (LPS) was derived from Escherichia coli 055:B5 (Difco, Detroit, MI). NO₂ was measured using a commercial Griess-Romijn Chemical, (Wako Pure reagent Tokyo, Japan). Monochlorobimane (MCB) was from Molecular Probes (Eugene, OR). Mouse IFN-γ was from R & D Systems (Minneapolis, MN). Mouse IL-10 was from Genzyme (Cambridge, MA). Human IL-12, anti-CD4 antibody and anti-CD3 antibody were from PharMingen (San Diego, CA). Human IL-2, IL-6 and lentinan [a β -(1–3)-glucan from edible mushroom, Lentinus edodes Berk. (Sing)] are the products of Ajinomoto (Tokyo, Japan).

Peritoneal macrophages

The procedures were published previously (27). Briefly, peritoneal cells were harvested by injecting 5 ml of an icecooled phenol red-free RPMI 1640 medium (Nikken Seibutsu, Kyoto, Japan) without antibiotics into the peritoneal cavity. The collected cells from five or more mice were cultured in a microplate (167008; Nunc, Roskilde, Denmark) at $1-3 \times 10^5/$ 200 µl in the same medium. Cells were plated in quadruplicate wells. The adherent cells were used as resident peritoneal macrophages. RMp or OMp were induced by administering 20 mg of GSH-OEt or 200 µg of (NACOMe)₂ i.p. 1 day before the collection. Macrophages were stimulated with 100 U/ml IFN- γ and/or 100 ng/ml LPS for cytokine and NO production for 48 h.

Nitrite and cytokine determination

Briefly, cell-free supernatants were incubated with Griess reagent for 10 min and absorbance at 550 nm was measured using an Arvo multilabel counter (Wallac, Turuk, Finland). The cytokine concentration was determined using an ELISA kit (Opt EIA; PharMingen) according to the manufacturer's instructions.

Qualitative determination of intracellular GSH with ACAS

First, 300 μ l of peritoneal cells suspension adjusted to a cell density of 3 \times 10⁵ cells/ml medium was charged into a Lab-Tek chamber slide (136439; Nunc) and incubated for 3 h. After washing, 300 μ l of 10 μ M MCB was added and incubated for 30 min. Then the fluorescent intensity was monitored by argonion laser cytometry with a ACAS 570 work station (Meridian, Okemos, MI). Intracellular GSH levels were detected with excitation and emission wavelengths of 350 and 460 nm.

CD4⁺ T cell culture and T_h1 polarization assay

Spleens from five to 10 mice were pooled, and CD8⁺ T cells and B cells were first removed using magnetic microbeads (Polyscience, Warrington, PA) coated with anti-CD8 antibody (53-6.7) and anti-HSA antibody (M16/9). CD4⁺ T cells were further positively selected with an auto-MACS (Miltenyi Biotech, Bergisch-Gladbach, Germany) using microbeads



Fig. 3. Intracellular GSH content of RMp and OMp. The amount of intracellular GSH in adherent macrophages of the mice indicated (n = 5) was monitored by argon-ion laser cytometry. The yellowish to red or blue color indicates the presence of abundant intracellular GSH. Alternatively IL-2R γ^{γ} mice were injected i.p. with 20 µg (NACOMe)₂ 24 h before the collection of macrophages. Results shown are representative of two separate experiments.

coated with anti-CD4 antibody. The purity of the fractionated CD4⁺ T cells was consistently confirmed to be >99% purity using FACScan (Becton Dickinson, Franklin Lakes, NJ). Macrophages were added at a cell density of 1 × 10⁵/well onto the microplates (Becton Dickinson) pre-coated with anti-CD3 antibody, adhered for 3 h followed by washing and were then co-cultured together with purified CD4⁺ T cells (5 × 10⁵/ well) for 48 h.

Statistical analysis

Results are expressed as means \pm SE. Significant differences were calculated using Student's *t*-test. *P* < 0.05 was considered statistically significant.

Results

Life longevity and the onset of IBD-like diseases in IL-2R $\gamma^{/-}$ and JAK3-/- mice

The mortality and the extent of colitis were investigated in IL-2R γ or JAK3 gene-disrupted mice. A significant reduction in survival rate was observed (IL-2R^{-/Y} mice 2.6 months and JAK3^{-/-} male mice 8.8 months, Fig. 1a) compared with normal C57BL/6 male mice (21.5 months). Whereas the wild-type littermates never manifested the IBD-like symptoms spontaneously, both IL-2R $\gamma^{\prime/-}$ and IL-2R $\gamma^{\prime/-}$ spontaneously displayed the symptoms within 4 months. Half of the IL-2R γ^{-} mice exhibited anal prolapse by 12 weeks and JAK3^{-/-} (male) by 26 weeks (Fig. 1b). The features of colitis in IL-2R $\gamma^{-/-}$, IL-2R $\gamma^{-/-}$ and JAK3-/- were characterized by intestinal shortening, colonal hypertrophy, diarrhea, loose passage, bloody stool and anal prolapse. Pathological analysis was conducted on specimens of the large intestine between 2 and 3 cm proximal to the anus. The colon was opened longitudinally and fixed in 4% formaldehyde. After embedding in paraffin, 4-um serial sections were prepared and stained with hematoxylin. Neither infiltration of inflammatory cells nor aberrant changes in goblet cells and mucosal epithelium cells were observed in wild-type. In contrast, the hyperplasia of the mucosal epithelium was 2-3 times thicker than those of wild-type, and the shortening of colon length became evident in IL-2R $\gamma^{-/-}$ and IL-2R $\gamma^{-/Y}$ by 13 weeks. Similar changes were also detected in JAK3-/- by 24 weeks, although the disease severity indexed by macroscopic inflammation score was milder than that in IL-2R $\gamma^{-/-}$ and IL- $2R\gamma^{N}$ (Fig. 1c). Infiltration of macrophages and lymphocytes as a local cluster was observed only in the lamina propria mucosa, and the infiltration of neutrophils was not observed. The cluster was only observed at a low level in the bottom of



Fig. 4. Skewing to RMp phenotypes in γ_c gene-disrupted mice. (a) Macrophages collected from IL-2R $\gamma^{+/-}$ or IL-2R $\gamma^{-/-}$ mice at the age of 10 weeks were evaluated for their NO-producing capability, IL-12 and IL-10. RMp were induced *in vitro* by treating with 100 mM GSH-OEt 24 h. macrophages were stimulated for 48 h with 100 U/mI IFN- γ and 100 ng/mI LPS for IL-12 and NO production, and with 100 ng/mI LPS for IL-10. (b) IL-2R $\gamma^{+/\gamma}$ (23 weeks old) and IL-2R $\gamma^{-/\gamma}$ mice with anal prolapse (17–23 weeks old) were analyzed. The procedures were the same as those in (a). IFN- γ production was also confirmed with a culture stimulated with 100 ng/mI LPS. Macrophages were collected from at least six mice for each experiment. Results shown are representative of two separate experiments.

the lamina propria mucosa (Fig. 2). When autopsies were conducted immediately before the occurrence of anal prolapse, crypt distortion, degeneration and loss of goblet cells, and metaplasia and degeneration of mucosal epithelium were identified at high levels (Fig. 2).

Periodical change of thiol redox status in macrophages at the early stage of IBD

Interestingly, macrophages of IL-2R γ^{-} of 6 weeks elicited functional phenotypes of OMp with reduced NO, IL-12 and IFN- γ , and elevated IL-10 production (data not shown), and intracellular GSH (icGSH) was reduced compared with those of wild-type and IL-2R γ^{-} (Fig. 3c and d versus a and b). However, the OMp phenotypes varied later to those of RMp with increased icGSH at the age of 10 weeks (Fig. 3e) and the functional phenotypes became comparable to macrophages

Pathogenesis of IBD and thiol redox status of macrophages 631

of IL-2R $\gamma^{/-}$ treated with GSH-OEt (Fig. 4a). The conversion to RMp phenotypes was functionally evident also for IL-2R $\gamma^{/\gamma}$ of 13 weeks, with the anal prolapse (Fig. 4b).

Conversion of RMp to OMp and a prophylactic intervention

RMp of IL-2R γ^{-} of 10 weeks could be converted to OMp by *in vitro* treatment with 1 mM (NACOMe)₂ for 24 h (Fig. 3f). (NACOMe)₂ is a cystine derivative which efficiently reduces the intracellular GSH content in macrophages and is possible to apply in the clinic (27,28). Thus, converted OMp elicited a reduced potential to produce IL-12 and NO (Fig. 5a and b). The conversion to OMp by i.p. administration of 20 µg (NACOMe)₂ (5 times, twice a weeks starting at the age of 9.5 weeks) delayed the onset of anal prolapse and increased the survival rate (Fig. 5c and d). This indicates that the blockade of conversion from OMp to RMp by (NACOMe)₂ may interfere with the occurrence of IBD-like symptoms, which was also confirmed by histological studies on specimens of the large intestine.

Conversion of T_h2 skewing to T_h1 , concomitant with the onset of IBD symptoms

As expected, the IL-2R $\gamma^{-\gamma}$ that survived for 23 weeks and JAK3-/- for 14 weeks without IBD-like symptoms maintained the phenotypes of OMp with reduced NO, IL-12 and IFN-γ, and elevated IL-10 production (data not shown). Consistent with this observation, the $T_h 1/T_h 2$ balance was skewed to $T_h 2$ in these OMp prone mice, and they also showed lowered IFNy:IL-4 and IFN-y:IL-10 ratios, which was deduced from the amount of cytokines produced by purified CD4⁺ T cells (Fig. 6a and b). Considering the late onset of IBD symptoms in JAK3^{-/-} (Fig. 1b), the skewing to T_h1 was expected to be later than in IL-2Ry gene-disrupted mice. Accordingly the skewing became evident initially at the age of 5.5 months (Fig. 7c), when the occurrence of the anal prolapse was detected in 40% of JAK3-/- (Fig. 1b). In vivo application of GSH-OEt converted OMp in JAK3-/- of 20 weeks to RMp and endowed the potential to produce IL-12 (Fig. 8a and b). GSH-OEt is a cell-permeable GSH derivative which efficiently induces RMp in vivo (27,28). In parallel with this conversion, GSH-OEt accelerated the onset of the IBD of OMp prone JAK3^{-/-} leading to 90% mortality in 4 months when injected 3 times a week for 2 weeks from 20 to 22 weeks (Fig. 8c). A β -(1–3)-glucan, lentinan, a potent inducer of RMp/T_h1 (24,28), structurally far removed from GSH-OEt, also accelerated the onset comparable to GSH-OEt when injected at a dose of 20 µg on a similar schedule.

T_{h2} cytokines augmented the effector function of macrophages depending on the redox status

The impact of T_h2 cytokines on the function of OMp was investigated to provide a preliminary clarification of the mechanism underlying the conversion of OMp/T_h2 to RMp/T_h1. T_h2 cytokines, IL-4, IL10 and IL-13, are well known to be anti-inflammatory or immunosuppressive cytokines, and the action is mainly mediated by the suppressive effect on the effector functions of macrophages, including the reduction of the potential to produce NO. Correspondingly, macrophages of JAK3^{+/+} or IL-2Rγ^{+/Y} elicited a reduced potential to produce NO when exposed to either of IL-4, IL10 or IL-13 for 24 h, and

632 Pathogenesis of IBD and thiol redox status of macrophages



Fig. 5. Conversion of RMp to OMp and a prophylactic intervention. RMp of $IL-2R\gamma^{-1}$ of 10 weeks of age were skewed to OMp by *in vitro* treatment with 1 mM (NACOMe)₂ for 24 h, and the culture was stimulated with LPS + IFN- γ to measure the produced NO (a) and IL-12 (b), as well as the production by non-treated or 10 mM GSH-OEt-treated macrophages [a and b; P < 0.01; medium versus (NACOMe)₂]. Results shown are representative of two separate experiments. The skewing to OMp by i.p. administration of 20 µg (NACOMe)₂ (5 times, twice a week starting at the age of 9.5 weeks) delayed the onset of anus prolapse (c, n = 12) and increased the survival rate (d, n = 12).



Fig. 6. The skewing of the $T_h 1/T_h 2$ balance in IL-2R γ gene-disrupted mice. CD4⁺ T cells were purified from spleen cells of IL-2R $\gamma^{+/Y}$ and IL-2R $\gamma^{-/Y}$ mice surviving for 23–29 weeks without anal prolapse, and of JAK3^{+/+}, JAK3^{+/-} and JAK3^{-/-} mice between 12 and 16 weeks of age. After stimulation with anti-CD3 for 48 h the supernatants were assayed for IFN- γ , IL-4 and IL-10, and IFN- γ :IL-4 (a) and IFN- γ :IL-10 (b) ratios were deduced. The amounts of secreted cytokines are also indicated (c–e). Results shown are representative of two separate experiments.

the extent of reduction was comparable to transforming growth factor (TGF)- β exposure. On the contrary, however, macrophages of JAK3^{-/-} or IL-2R $\gamma^{-/Y}$ (OMp phenotype) behaved in a contrasting manner, elevating the NO production

potential (Fig. 9a). These T_h2 cytokines share the potential to regulate bilaterally the thiol redox status of macrophages, depending on the pre-existing redox status (data not shown). OMp were converted to RMp with either IL-4, IL10 or IL-13,



Fig. 7. The skewing of the $T_h 1/T_h 2$ balance in JAK3 gene-disrupted mice. CD4+ T cells were purified from spleen cells of JAK3^{+/-} and JAK3^{-/-} mice between 2 and 10.5 months of age were investigated (c: P < 0.01; 2, 4, 5.5 versus 10.5 month). IFN- γ (a), IL-4 (b) and IFN- γ :IL-4 ratio (c) were deduced as shown in Fig. 5. Results shown are representative of two separate experiments.

while RMp were converted to OMp, whereas TGF- β consistently induced OMp, irrespective of the pre-existing redox status.

Discussion

The spontaneous onsets of IBD-like diseases in IL-2Ry chain (γ_c) -deficient mice with a truncated mutation (Fig. 2) coincides well with the reported observation describing inflammatory changes in the intestine from 8 weeks of age onwards (39). JAK3 is an essential transducer of γ_c -dependent signals, so it is not surprising that JAK3^{-/-} elicited similar symptoms to IL- $2R\gamma^{/-}$ and IL- $2R\gamma^{//}$. It is of note that the T_h1/T_h2 balance was skewed to T_b2 (Figs 6 and 7) and macrophages to OMp in IL- $2R\gamma^{-/-}$, IL- $2R\gamma^{-/Y}$ and JAK3^{-/-} at an early stage of disease. The skewing of macrophages to OMp may be due to the impact of luminal antigens on the intracellular redox status of macrophages, although the pathological meaning of the skewing to OMp at an early stage of disease remains unclear. One possibility is the forced infiltration of macrophages into intestinal mucosa by modulating the chemotaxis of macrophages. A certain subset of macrophages selectively produces CC chemokines (40) and the thiol redox status affects the expression of adhesion molecules critical for transendothelial migration of the inflammatory cells (41).

IL-12 and IL-10 play a pivotal role in maintaining the *in vivo* balance between the T_h1 and T_h2 response (42–45). Concurrent with the conversion to RMp, IL-12 produced by RMp may participate in the disease progression through the induction of T_h1 skewing. T_h1 -type inflammation has a more pivotal role in CD, whereas T_h2 -type responses are more pivotal in UC, although the relative relevance of both responses is still controversial (7,8,46–50). Our findings that OMp/ T_h2 skewing shifted to RMp/ T_h1 skewing along with

disease progression (Fig. 4) and that (NACOMe)₂ prevented the onset of IBD-like symptoms (Fig. 5), together with the accelerated disease progression by RMp/Th1 skewing by GSH-OEt or a β -(1–3)-glucan (Fig. 8c), support the concept that T_h1-type inflammation has a relevant role in the later pathological progression of colitis, at least in our models. Initial skewing to OMp/Th2 prior to later conversion to RMp/Th1 is the pre-requisite for disease onset, because RMp/T_h1 skewing by GSH-OEt or a β -(1–3)-glucan in wild-type did not cause any sign of colitis at all (data not shown). The combined affect of genetic factors and luminal antigens on the redox status of macrophages may trigger the initial priming to OMp/T_b2. RMp may also participate in the disease progression through the production of NO by RMp (Fig. 4). It is becoming increasingly apparent that chronic inflammation in IBD is associated with the sustained overproduction of NO (51,52). The mechanism underlying the observed conversion of OMp/T_h2 to RMp/T_h1 is unclear. In this context it is of interest that Th2 cytokines elicited a bilateral effect to convert OMp to RMp and augmented NO production (Fig. 9). Th2 cytokine's function as either an anti-inflammatory or pro-inflammatory mediator may depend on the redox status of macrophages. Infiltration of inflammatory cells in inflamed sites may be triggered initially by OMp, resulting in the release of anti-inflammatory cytokines, which in turn may trigger the conversion of OMp to RMp. Subsequent tissue injury is ascribed to RMp. As such, depending on the redox status of macrophages, anti-inflammatory cytokines may elicit a paradoxical pro-inflammatory action. The sequential change of the redox status of macrophages has also turned out to be responsible for other autoimmune diseases, such as type I diabetes and hepatitis (Y. Murata, in preparation). The bilateral functions of T_b2 cytokines have also been reported (34). The role of the intracellular thiol redox status of macrophages in regulating

634 Pathogenesis of IBD and thiol redox status of macrophages



Fig. 8. Conversion of OMp to RMp accelerated the onset of IBD symptoms. *In vivo* application of GSH-OEt (i.p., 20 mg) converted OMp in JAK3^{-/-} of 20 weeks to RMp (a) and the macrophages collected were investigated for their potential to produce IL-12 (b, P < 0.01) according to the procedures described. GSH-OEt injected 3 times a week for 2 weeks from 20 to 22 weeks and lentinan injected at a dosage of 1 mg/kg at the same schedule accelerated the onset of IBD of JAK3^{-/-} (c, n = 10).



Fig. 9. $T_h 2$ cytokines augmented NO production by OMp. Macrophages of JAK3^{+/+}, JAK3^{-/-}, IL-2R $\gamma^{+/Y}$ or IL-2R $\gamma^{+/Y}$ mice were exposed to either IL-4 (10 ng/ml), IL-10 (50 ng/ml), IL-13 (10 ng/ml), TGF- β (20 ng/ml) or IL-2 (1000 U/ml) for 20 h, and then stimulated with 100 U/ml IFN- γ and 100 ng/ml LPS for 48 h to measure the production of NO. Results shown are representative of two separate experiments. **P* < 0.01 for medium control.

functional phenotypes of macrophages producing either NO, IFN-γ and IL-12 or IL-10 has also been proposed (24,26–28). The intestinal epithelium is likely to be exposed to oxidants derived from bacterial constituents or endogenously generated ROS by infiltrating macrophages and neutrophils within the lamina propria. GSH, being the major intracellular thiol, provides protection against oxidative injury (53). Clinical and experimental studies revealed that intestinal mucosa showed deficiency in GSH content. Accordingly, *in vivo* administration of *N*-acetyl-L-cysteine attenuates the acute colitis through increasing mucosal GSH levels, suggesting that GSH precursors may be of relevance in the acute relapse of IBD (54,55). Further study is required to define the interplay between OMp or RMp and mucosal epithelium, in the context of the influence of icGSH on the counteraction of infiltrating cells and intestinal mucosa. Lamina propria macrophages lack the capacity to elaborate cysteine and thereby secure the physiological unresponsiveness of lamina propria T cells to nominal antigen exposure (56). The local recruitment of blood monocytes in IBD may endow hyper-responsiveness with T cells (56). The proposed hypothesis may well be compatible with our notion that the conversion of OMp to RMp might underlie the hyperresponsiveness of T cells in IBD.

The most crucial and urgent issue to address is the analysis of the redox status of locally recruited macrophages into inflamed sites of the intestinal tract. GSH is present in virtually all cells, which makes it difficult to demonstrate directly the frequency of RMp/OMp in small and large intestines. The procedures needed for multi-step purification of locally recruited macrophages also accompany the artificial changes of redox status of macrophages due to the plasticity (conversion) of the redox status. To explore the relevant issue to compare the macrophages isolated from disease and normal sites, further elaboration of the new technology to define directly the content of GSH will be necessary. The further study will be needed to define the biological significance of the new insight for the potential role of RMp/OMp in the development of IBD. In this context the cytokine propensity of the intestinal T cells should be also analyzed in relation to the disease progression. Finally, it should be noted that the up-regulation of TLR4 in both UC and CD (57) indicates the necessity of studies on the influence of the redox status of macrophages on the TLR-MyD88-NF-kB system, considering the recruitment of macrophages to intestinal mucosa, accessible to luminal bacterial constituents.

Acknowledgements

The authors thank Drs S. Taki and T. Arase (Chiba University Medical School), H. Fujiwara (Osaka University Medical School), S. Koyasu and T. Ohteki (Keio University Medical School), and H. Dvorak (Harvard University) for their continuous encouragement and helpful advice, and Ms Yoko Hamuro and Naoko leda for secretarial assistance.

Abbreviations

CD	Crohn's disease
GSH	glutathione
GSH-OEt	glutathione monoethylester
GSSG	oxidized glutathione
IL-2R	IL-2 receptor
JAK	Janus family tyrosine kinase
icGSH	intracellular GSH
LPS	lipopolysaccharide
MCB	monochlorobimane
ОМр	oxidative macrophages
RMp	reductive macrophages
ROS	reactive oxygen species
TGF	transforming growth factor
TLR	Toll-like receptor
UC	ulcerative colitis

References

- 1 Fiocchi, C. 1998. Inflammatory bowel disease: etiology and pathogenesis. *Gastroenterology* 115:182.
- 2 Duchmann, R., Schmitt, E., Knolle, P., Meyer zum Buschenfelde, K. H. and Neurath, M. 1996. Tolerance towards resident intestinal flora in mice is abrogated in experimental colitis and restored by treatment with interleukin-10 or antibodies to interleukin-12. *Eur. J. Immunol.* 26:934.
- 3 Morales, V. M., Snapper, S. B. and Blumberg, R. S. 1996. Probing the gastrointestinal immune function using transgenic and knockout technology. *Curr. Opin. Gastroenterol.* 12:577.
- 4 Braegger, C. P. and MacDonald, T. T. 1994. Immune mechanisms in chronic inflammatory bowel disease. *Ann. Allergy* 72:135.
- 5 Brandtzaeg, P., Haraldsen, G. and Rugtveit, J. 1997. Immunopathology of human inflammatory bowel disease. *Semin. Immunopathol.* 18:555.
- 6 Dohi, T., Fujihashi, K., Kiyono, H., Elson, C. O. and Mcghee, J. R. 2000. Mice deficient in Th1- and Th2-type cytokines develop

Pathogenesis of IBD and thiol redox status of macrophages 635

distinct forms of hapten-induced colitis. *Gastroenterology* 119:724.

- 7 Breese, E., Braegger, C. P., Corrigan, C. J., Walker-Smith, J. A. and MacDonald, T. T. 1993. Interleukin-2- and interferon-gammasecreting T cells in normal and diseased human intestinal mucosa. *Immunology* 78:127.
- 8 Fuss, I. J., Neurath, M., Boirivant, M., Klein, J. S., de la Motte, C., Strong, S. A., Fiocchi, C. and Strober, W. 1996. Disparate CD4+ lamina propria (LP) lymphokine secretion profiles in inflammatory bowel disease. Crohn's disease LP cells manifest increased secretion of IFN-gamma, whereas ulcerative colitis LP cells manifest increased secretion of IL-5. *J. Immunol.* 157:1261.
- 9 Parronchi, P., Romagnani, P., Annunziato, F., Sampognaro, S., Becchio, A., Giannarini, L., Maggi, E., Pupilli, C., Tonelli, F. and Romagnani, S. 1997. Type 1 T-helper cell predominance and interleukin-12 expression in the gut of patients with Crohn's disease. Am. J. Pathol. 150:823.
- 10 Ma, A., Datta, M., Margosian, E., Chen, J. and Horak, I. 1995. T cells, but not B cells, are required for bowel inflammation in interleukin 2-deficient mice. *J. Exp. Med.* 182:1567.
- 11 Mackaness, G. B. 1964. The immunological basis of acquired cellular resistance. J. Exp. Med. 120:105.
- 12 Adams, D. O. and Hamilton, T. A. 1984. The cell biology of macrophage activation. *Annu. Rev. Immunol.* 2:283.
- 13 Means, T. K., Golenbock, D. T. and Fenton, M. J. 2000. Structure and function of Toll-like receptor proteins. *Life Sci.* 68:241.
- 14 Kaisho, T. and Akira, S. 2000. Critical roles of Toll-like receptors in host defense. *Crit. Rev. Immunol.* 20:393.
- 15 Muzio, M., Polentarutti, N., Bosisio, D., Manoj Kumar, P. P. and Mantovani, A. 2000. Toll-like receptor family and signaling pathway. *Biochem. Soc. Trans* 28:563.
- 16 Mahida, Y. R., Wu, K. C. and Jewell, D. P. 1989. Respiratory burst activity of intestinal macrophages in normal and inflammatory bowel disease. *Gut* 30:1362.
- 17 Williams, J. G. 1990. Phagocytes, toxic oxygen metabolites and inflammatory bowel disease: implications for treatment. *Ann. R. Coll. Surg. Engl.* 72:253.
- 18 Grisham, M. B. and Granger, D. N. 1988. Neutrophil-mediated mucosal injury. Role of reactive oxygen metabolites. *Dig. Dis. Sci.* 33:6S.
- 19 Weiss, S. J. 1989. Tissue destruction by neutrophils. *N. Engl. J. Med.* 320:365.
- 20 Grisham, M. B., MacDermott, R. P. and Deitch, E. A. 1990. Oxidant defense mechanisms in the human colon. *Inflammation* 6:669.
- 21 Meister, A. 1989. Glutathione Centennial; Molecular Properties and Clinical Applications, Academic Press, New York.
- 22 Grisham, M. B. 1994. Oxidants and free radicals in inflammatory bowel disease. *Lancet* 344:859.
- 23 Gross, V., Arndt, H. and Andus, T., Palitzsch, K. D. and Scholmerich, J. 1994. Free radicals in inflammatory bowel diseases. pathophysiology and therapeutic implications. *Hepatogastroenterology* 41:320.
- 24 Hamuro, J., Murata, Y. and Suzuki, M. 1999. The triggering and healing of tumor stromal inflammatory reactions regulated by oxidative and reductive macrophages. *Gann. Monogr. Cancer Res.* 48:153.
- 25 Peterson, J. D., Herzenberg, L. A., Vasquez, K. and Waltenbaugh, C. 1998. Glutathione levels in antigen-presenting cells modulate T_h1 versus T_h2 response patterns. *Proc. Natl Acad. Sci. USA* 95:3071.
- 26 Murata, Y., Amao, M., Yoneda, J. and Hamuro, J. 2002. Intracellular thiol redox status of macrophages directs the T_h1 skewing in thioredoxin transgenic mice during aging. *Mol. Immunol.* 38:747.
- 27 Murata, Y., Shimamura, T. and Hamuro, J. 2002. The polarization of $T_h 1/T_h 2$ balance is dependent on the intracellular thiol redox status of macrophages due to the distinctive cytokines production. *Int. Immunol.* 14:201.
- 28 Murata,Y., Shimamura, T., Tagami, T., Takatsuki, F. and Hamuro, J. 2002. The skewing to T_h1 induced by lentinan is directed through the distinctive cytokine production by macrophages with

636 Pathogenesis of IBD and thiol redox status of macrophages

elevated intracellular glutathione content. *Int. Immunopharmacol.* 2:673.

- 29 Utsugi, M., Dobashi, K., Koga, Y., Shimizu, Y., Ishizuka, T., Iizuka, K., Hamuro, J. Nakazawa, T. and Mori, M. 2002. Glutathione redox regulates lipopolysaccharide induced IL-12 production through p38 mitogen-activated protein kinase activation in human monocytes: role of glutathione redox in IFN-γ priming of IL-12 production. *J. Leukoc. Biol.* 71:339.
- 30 Dobashi, K., Aihara, M., Araki, T., Shimizu, Y., Utsugi, M., Iizuka, K., Murata, Y., Hamuro, J., Nakazawa, T. and Mori, M. 2001. Regulation of LPS induced IL-12 production by IFN-γ and IL-4 through intracellular glutathione status in human alveolar macrophages. *Clin. Exp. Immunol.* 124:290.
- 31 Nakamura, H., Nakamura, K. and Yodoi, J. 1997. Redox regulation of cellular activation. Annu. Rev. Immunol. 15:351.
- 32 Suzuki, Y. J., Mizuno, M. and Packer, L. 1994. Signal transduction for nuclear factor-kappa B activation. Proposed location of antioxidant-inhibitable step. *J. Immunol.* 153:5008.
- 33 Munder, M., Eichmann, K. and Modolell, M. 1998. Alternative metabolic states in murine macrophages reflected by the nitric oxide synthase/arginase balance: competitive regulation by CD4+ T cells correlates with T_h1/T_h2 phenotype. *J. Immunol.* 160:5347.
- 34 Goerdt, S. and Orfanos, C. E. 1999. Other functions, other genes: alternative activation of antigen-presenting cells. *Immunity* 10:137.
- 35 Shearer, J. D., Richards, J. R., Mills, C. D. and Caldwell, M. D. 1997. Differential regulation of macrophage arginine metabolism: a proposed role in wound healing. *Am. J. Physiol.* 272:E181.
- 36 Mills, C. D., Kincaid, K., Alt, J. M., Heilman, M. J. and Hill, A. M. 2000, M-1/M-2 macrophages and the T_h1/T_h2 paradigm. *J. Immunol.* 164:6166.
- 37 Park, S. Y., Saijo, K., Takahashi, T., Osawa, M., Arase, H., Hirayama, N., Miyake, K., Nakauchi, H., Shirasawa, T. and Saito, T. 1995. Developmental defects of lymphoid cells in JAK3 kinasedeficient mice. *Immunity* 3:771.
- 38 Ohbo, K., Suda, T., Hashiyama, M., Mantani, A., Ikebe, M., Miyakawa, K., Moriyama, M., Nakamura, M., Katsuki, M., Takahashi, K., Yamamura, K. and Sugamura, K. 1996. Modulation of hematopoiesis in mice with a truncated mutant of the interleukin-2 receptor gamma chain. *Blood* 87:956.
- 39 Ikebe, M., Miyakawa, K., Takahashi, K., Ohbo, K., Nakamura, M., Sugamura, K., Suda, T., Yamamura, K. and Tomita, K. 1997. Lymphohaematopoietic abnormalities and systemic lymphoproliferative disorder in interleukin-2 receptor gamma chain-deficient mice. Int. J. Exp. Pathol. 78:133.
- 40 Kodelja, V., Mueller, C., Politz, O., Hakij, N., Orfanos, C. E. and Goerdt, S. 1998. Alternative macrophage activation-associated CC-chemokine-1, a novel structural homologue of macrophage inflammatory protein-1α with a T_h2-associated expression pattern. *J. Immunol.* 160:1411.
- 41 Kokura, S., Robert, E., Wolf, T., Yoshikawa, T. D., Granger, N. and Aw, T. Y. 1999. Molecular mechanisms of neutrophil–endothelial cell adhesion induced by redox imbalance. *Circ. Res.* 84:516.
- 42 Chehimi, J. and Trinchieri, G. 1994. Interleukin-12: a bridge between innate resistance and adaptive immunity with a role in infection and acquired immunodeficiency. *J. Clin. Immunol.* 14:149.

- 43 Hsieh, C. S., Heimberger, A. B., Gold, J. S., O'Garra, A. and Murphy, K. M. 1992. Differential regulation of T helper phenotype development by interleukins 4 and 10 in an alpha beta T-cellreceptor transgenic system. *Proc. Natl Acad. Sci. USA* 89:6065.
- 44 Powrie, F., Leach, M. W., Mauze, S., Menon, S., Caddle, L. B. and Coffman, R. L. 1994. Inhibition of T_h1 responses prevents inflammatory bowel disease in scid mice reconstituted with CD45RB^{hi} CD4⁺ T cells. *Immunity* 1:553.
- 45 Rennick, D. M., Fort, M. M. and Davidson, N. J. 1997. Studies with IL-10-/- mice: an overview. J. Leuk. Biol. 61:389.
- 46 Neurath, M. F., Fuss, I., Kelsall, B. L., Stuber, E. and Strober, W. 1995. Antibodies to interleukin-12 abrogate established experimental colitis in mice. *J. Exp. Med.* 182:1281.
- 47 Elson, C. O., Beagley, K. W., Sharmanov, A. T., Fujihashi, K., Kiyono, H., Tennyson, G. S., Cong, Y., Black, C. A., Ridwan, B. W. and McGhee, J. R. 1996. Hapten-induced model of murine IBD: mucosal immune response and protection by tolerance. *J. Immunol.* 157:2174.
- 48 Strober, W., Kelsall, B., Fuss, I., Marth, T., Ludviksson, B., Ehrhardt, R. and Neurath, M. 1997. Reciprocal IFN- γ and TGF- β responses regulate the occurrence of mucosal inflammation. *Immunol. Today* 18:61.
- 49 Niessner, M. and Vold, B. A. 1995. Altered T_h1/T_h2 cytokine profiles in the intestinal mucosa of patients with inflammatory bowel disease as assessed by quantitative reversed transcribed polymerase chain reaction (RT-PCR). *Clin. Exp. Immunol.* 101:428.
- 50 Monteleone, G., Biancone, L., Marasco, R., Morrone, G., Marasco, O., Luzza, R. and Pallone, F. 1997. Interleukin 12 is expressed and actively released by Crohn's disease intestinal lamina propria mononuclear cells. *Gastroenterology*. 112:1169.
- 51 Grisham, M. B., Specian, R. D. and Zimmerman, T. E. 1994. Effects of nitric oxide synthase inhibition on the pathophysiology observed in a model of chronic granulomatous colitis. *J. Pharmacol. Exp. Ther.* 271:1114.
- 52 Aiko, S. and Grisham, M. B. 1995. Spontaneous intestinal inflammation and nitric oxide metabolism in HLA-B27 transgenic rats. *Gastroenterology* 109:142.
- 53 Benard, O. and Balasubramanian, K. A. 1993. Effect of oxidant exposure on thiol status in the intestinal mucosa. *Biochem. Pharmacol.* 45:2011.
- 54 Ardite, E., Sans, M., Panes, J., Romero, F. J., Pique, J. M. and Fernandez-Checa, J. C. 2000. Replenishment of glutathione levels improves mucosal function in experimental acute colitis. *Lab. Invest.* 80:735.
- 55 Nieto, N., Torres, M. I., Fernandez, M. I., Giron, M. D., Rios, A., Suarez, M. D. and Gil, A. 2000. Experimental ulcerative colitis impairs antioxidant defense system in rat intestine. *Dig. Dis. Sci.* 45:1820.
- 56 Sido, B., Braunstein, J., Breitkreutz, R., Herfarth, C. and Meuer, S. C. 2000. Thiol-mediated redox regulation of intestinal lamina propria T lymphocytes. *J. Exp. Med.* 192:907.
- 57 Cario, E. and Podolsky, D. K. 2000. Differential alteration in intestinal epithelial cell expression of toll-like receptor 3 (TLR3) and TLR4 in inflammatory bowel disease. *Infect. Immun.* 68:7010.